

United States Department of Agriculture
Grain Inspection, Packers and Stockyards Administration
Federal Grain Inspection Service

FGIS Issuance Change

CHANGE TO☐ DIRECTIVE☐ MANUAL☒ HANDBOOK**CHANGE NO:**
10**TO (No.)****TITLE:**
Aflatoxin Handbook**DATE:**
1-31-05

PURPOSE OF CHANGE: The Aflatoxin Handbook has been revised to include instructions for the RIDASCREEN® Fast Aflatoxin SC, and the QuickTox SC test kits, amend the name of the RIDASCREEN® Fast Aflatoxin test method to RIDASCREEN® Fast Aflatoxin Total, and to make minor editorial changes to chapter 1.

FILING INSTRUCTIONS

Remove	Dated	Insert	Dated
Table of Contents	1-5-04	Table of Contents	1-31-05
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Chapter 6	1-5-04	Chapter 6	1-31-05
*****	*****	Chapter 12	1-31-05
*****	*****	Chapter 13	1-31-05

Retain this issuance sheet as an aid in verifying handbook contents.

/s/ David Orr

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Distribution: A, C, E

Originating Office: PPB, FMD

U.S. DEPARTMENT OF AGRICULTURE
GRAIN INSPECTION, PACKERS AND STOCKYARDS
ADMINISTRATION
FEDERAL GRAIN INSPECTION SERVICE
STOP 3630
WASHINGTON, D.C. 20090-3630

AFLATOXIN HANDBOOK
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1-31-05

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1.5 APPROVED TEST METHODS

FGIS has approved test kits for use at field testing locations. The AflaCup, and Agri-Screen and QuickTox test kits are approved for qualitative analysis of corn. The Aflatest, Fluoroquant, Veratox-AST, Myco✓, RIDASCREEN Fast Aflatoxin Total and RIDASCREEN Fast Aflatoxin SC test kits provide quantitative analysis but can be used for qualitative results. High Performance Liquid Chromatography (HPLC) testing is reserved for quantitative testing at the Technical Services Division (TSD) only.

FGIS APPROVED TEST METHODS			
Method and Test Kit	Approved for		Test Kit Range
	Qualitative	Quantitative	
QuickTox - (Envirologix)	X		20 ppb
AflaCup (International Diagnostics Inc.)	X		20 ppb
AgriScreen - (Neogen)	X		20 ppb
Veratox AST - (Neogen)	X	X	5 - 300 ppb (quantitative)
Fluoroquant - (Romer)	X	X	5 - 300 ppb (quantitative)
Aflatest – (Vicam)	X	X	5 - 300 ppb (quantitative)
Myco✓ - (Strategic Diagnostics Inc.)	X	X	5 - 80 ppb (quantitative)
RIDASCREEN Fast Aflatoxin Total (r-Biopharm)	X	X	5 - 50 ppb (quantitative)
RIDASCREEN Fast Aflatoxin SC (r-Biopharm)	X	X	5 - 100 ppb (quantitative)

NOTE: The test ranges are for performing an individual analysis with an undiluted sample extract. To obtain accurate results above the test kit range a supplemental analysis must be performed.

Listed in the table below are the test kits that are commonly used for official aflatoxin analysis. Use the table to determine the appropriate test kit(s) to use for testing the listed grain/commodity. For information concerning the testing of mixed grain, contact the Policies and Procedures Branch.

GRAIN/ COMMODITY	TEST METHOD								
	AflaCup	Aflatest	Agri-Screen	Fluoroquant	Myco✓	QuickTox	Ridascreen Fast Aflatoxin Total	Ridascreen Fast Aflatoxin SC	Veratox- AST
Corn	X	X	X	X	X	X	X	X	X
Sorghum		X		X	X		X	X	X
Wheat		X		X			X	X	X
Soybeans		X		X			X	X	X
Corn Screenings		(*)							(*)
Corn Meal		X		X	X		X	X	X
Corn Germ Meal		X					X	X	X
Corn Gluten Meal		X					X	X	X
Corn/Soy Blend		X		X	X		X	X	X
Corn Gluten Feed		X							
Flaking Corn Grits		X		(*)					(*)
Corn Flour								X	(*)
Corn Bran									(*)
Popcorn		X		X	X		X	X	X
Milled Rice		X		X			X		X
Rough Rice									(*)
Cracked Corn	(*)	(*)	(*)	(*)	(*)		X	X	(*)

NOTE: An X entered into a block denotes that the test kit has been evaluated and approved for the grain/commodity.

The symbol (*) entered into a block denotes that the test kit is under evaluation by TSD for the grain/commodity and is temporarily approved for official use.

CHAPTER 6

RIDASCREEN® FAST AFLATOXIN TOTAL TEST KIT

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6.1 GENERAL INFORMATION

The RIDASCREEN® FAST Aflatoxin Total test is a competitive enzyme immunoassay for the quantitative analysis of aflatoxin in select grains and commodities. **The test kit is limited to providing aflatoxin measurements between 5 – 50 ppb.** Accurate aflatoxin measurements above 50 ppb can be obtained by performing a supplemental analysis involving a diluted extract.

6.2 PREPARATION OF EXTRACTION SOLUTION

The extraction solvent used in the RIDASCREEN® FAST Aflatoxin Total test is a methanol/water (distilled or deionized) mixture consisting of 70 percent methanol (ACS grade or better) and 30 percent water.

- a. Using a graduated cylinder, measure 700 ml of methanol and place it into a clean carboy with spigot.
- b. Add 300 ml deionized or distilled water to the methanol and shake vigorously until it is completely mixed.
- c. Label the container stating the mixture (70 percent methanol and 30 percent water), date of preparation, and initials of technician who prepared the solution.
- d. Store this solution at room temperature in a tightly closed container until needed.

NOTE: To prepare smaller or larger amounts of solution use the ratio of 7 parts methanol to 3 parts of deionized or distilled water.

6.3 EXTRACTION PROCEDURES

- a. Transfer 50 grams of ground sample into an extraction mixing jar.
- b. Add 250 ml of the (70/30) methanol/water extraction solvent.
- c. Cover the extraction jar and blend on high speed for 2 minutes.
- d. Filter the extract through a filtering syringe.
- e. Dilute 1 ml of the filtrate with 1 ml of distilled/deionized water.

6.4 TEST PROCEDURES

a. Sample Analysis.

- (1) Allow reagents, microwells, and sample extracts to reach room temperature prior to running the test.
- (2) Insert a sufficient number of wells into the microwell holder for all standards and samples to be tested. (For example: to test 11 samples use 16 wells - 5 for the standards and 11 for the test samples).

Test Strip #1

Well #	1	2	3	4	5	6	7	8
Sample	C 0	C 4	C 10	C 20	C 50	S1	S2	S3

Test Strip #2

Well #	1	2	3	4	5	6	7	8
Sample	S4	S5	S6	S7	S8	S9	S10	S11

Where C 0 is the zero control, C 4 is the 4 ppb control, C 10 is the 10 ppb control, C 20 is the 20 ppb control, and C 50 is the 50 ppb control. S1 is sample 1, S2 is sample 2, S3 is sample 3, etc.

NOTE: Do not run more than 3 strips (19 samples) per set of control standards.

- (3) Using a new pipette tip for each standard and sample, pipet 50 μ l of standards and prepared sample to separate wells.
- (4) Add 50 μ l of enzyme conjugate (red capped bottle) into each well.
- (5) Add 50 μ l of anti-aflatoxin antibody (black capped bottle) into each well.
- (6) Mix thoroughly by gently sliding the plate back and forth on a flat surface.

- (7) Incubate for 5 minutes (\pm 0.5 minutes) at room temperature.
- (8) Dump the contents of the wells. Turn the wells upside down and tap out on a paper towel until the remaining liquid has been removed.
- (9) Using a wash bottle, fill each well with distilled or deionized water. Empty the wells again and remove all remaining liquid. Repeat this step 2 times (total of 3 washes).
- (10) Add 100 μ l of substrate/chromagen (white dropper bottle) to each well.
- (11) Mix thoroughly by gently sliding the plate back and forth on a flat surface.
- (12) Incubate for 5 minutes (\pm 0.5 minutes) at room temperature (64 – 86° F). Cover the wells with a paper towel to protect them from light sources.
- (13) Add 100 μ l of stop solution (yellow or orange dropper bottle) to each well.
- (14) Mix thoroughly by gently sliding the plate back and forth on a flat surface.
- (15) Measure absorbance at 450 nm using the Biotek EL 301, or Awareness Technology Stat-Fax Model 303 PLUS microwell readers.

(Results must be read within 10 minutes)

b. Reading Results with the Microwell Reader.

- (1) Biotek EL 301 Microwell Reader.
 - (a) Make sure that the microwell reader is on and allowed to warm-up for a minimum of 15 minutes before using.
 - (b) Remove sample carriage and hit "Enter."
 - (c) Insert W2 filter and hit "Enter."
 - (d) Insert W1 filter (450 nm) and hit "Enter."
 - (e) Hit "Clear" and then "Blank." This will cause the instrument to read air as the blank sample.

- (f) Load microwells into sample carriage so that the first control labeled 0 is in position A1.
- (g) Load the sample carriage into the strip reader so that position A1 is under the light beam of the reader.
- (h) Press "Read" and an absorbance value for A1 should appear in the display on the microwell reader. Record the value.
- (i) Slide the carriage to position A2 and press "Read." An absorbance value for A2 will appear. Record the value.
- (j) Repeat step (i) until absorbance values have been obtained for all controls and samples. Record the values.
- (k) Use the RIDA®SOFT Win Data software provided by r-Biopharm to convert the absorbance values into concentration values.

(2) Stat-Fax Model 303 PLUS Microwell Reader

- (a) To begin from the "Ready" prompt, press Menu, key in the test number, and then press Enter.
- (b) The screen will read, "Set carrier to A, press enter." Place the wells all the way to the right in the carrier. Push the carrier all the way to the left to line up the notch with the wells, then press enter. The carrier will advance into the reader, and it should start to print.
- (c) When the reader is finished reading the strip, the screen will read, "Plot Curve Y/N?"

Press "Yes" (1/A) to print the graph,

Press "No" (0) to skip this feature.
- (d) The screen will read, "Accept Curve Y/N ?"

Press "Yes" (1/A) to accept the curve and proceed to read another strip. When finished reading the second strip, press "Clear" twice and the results strip will print, "Test Ended."

Press "No" (0) to end the test.

6.5 REPORTING AND CERTIFYING TEST RESULTS

- a. Report all results on the pan ticket and the inspection log to the nearest whole ppb.
- b. Sample results over 50 ppb are reported as >50 ppb unless a supplemental analysis is performed.
- c. Refer to the Certification section of the handbook for more detailed certification procedures.

6.6 SUPPLEMENTAL ANALYSIS

- a. Diluting the Sample Extract.

If quantitative results are above the testing limits (i.e., 50 ppb) of the test kit, test results are reported as exceeding the limit. To determine and report an aflatoxin level higher than 50 ppb, the sample extract must be diluted so that a value between 5 and 50 ppb is obtained.

The final aflatoxin concentration is calculated by multiplying the results with the diluted extract by the dilution factor.

- b. Example.

If the original analysis reported the aflatoxin value at 70 ppb, the sample extract would be diluted using the following procedures in order to obtain a true value.

- (1) Dilute 5 ml of the original extract with 5 ml of the extraction solvent mixture. The total volume is 10 ml. This is a 1 to 2 dilution (compares volume in the beginning with the total volume in the end).
- (2) Proceed to sample analysis .
- (3) Multiply the analytical results obtained by 2 to obtain the actual aflatoxin concentration. For example, if 34 ppb was the original value obtained with the diluted extract, the actual concentration in the original sample was 68 ppb.

$$\text{True Aflatoxin Value} = \frac{\text{Total Volume}}{\text{Initial Extract Volume}} \times \text{Aflatoxin Result}$$

$$\begin{aligned}\text{True Aflatoxin Value} &= (10 \div 5) \times 34 \text{ ppb} \\ &= 2 \times 34 \text{ ppb} = 68 \text{ ppb}\end{aligned}$$

6.7 CLEANING LABWARE

a. Negative Tests (≤ 20 ppb).

(1) Labware.

Prepare a solution consisting of dishwashing liquid and water. Completely submerge the used glassware, beakers, etc., wash thoroughly, then rinse with clean water before reusing.

(2) Disposable Materials.

Place materials in a garbage bag for routine trash disposal.

b. Positive Tests (> 20 ppb).

(1) Labware.

Prepare a bleach solution consisting of 1 part bleach to 10 parts water (e.g., 100 ml bleach to 1,000 ml water). Completely submerge the used glassware, beakers, etc., and soak for at least 5 minutes. Remove items from the bleach/water solution, submerge in a dishwashing liquid/water solution, wash thoroughly, then rinse with clean water before reusing.

(2) Disposable Materials.

Prepare a bleach solution consisting of 1 part bleach to 10 parts water in a plastic pail labeled "bleach solution". Soak disposable materials, such as used columns, cuvettes, vials, test kit components, etc., for at least 5 minutes. Pour the liquid down the drain and place the materials in a garbage bag and discard.

6.8 WASTE DISPOSAL

a. Negative Results (≤ 20 ppb).

If the test result is negative (equal to or less than 20 ppb), discard the syringe into a plastic garbage bag for disposal.

b. Positive Results (> 20 ppb).

If the result is positive (more than 20 ppb), the remaining ground portion must be decontaminated, using bleach, prior to disposal. Discard the filter syringe and remaining ground portion into a plastic garbage bag for disposal. The bleach rinse filtrate collected may be treated as a non-hazardous solution and disposed of by pouring down the drain.

6.9 EQUIPMENT AND SUPPLIES

a. Materials Supplied in Test Kits

- (1) 1 microtiter plate.
- (2) 48 antibody coated microwells.
- (3) 5 aflatoxin standard solutions of 1.3 ml each; 0, 4, 10, 20, and 50 ppb aflatoxins.
- (4) 1 red-capped bottle of 3 ml peroxidase conjugated aflatoxin solution.
- (5) 1 black-capped bottle of 3 ml anti-aflatoxin antibody.
- (6) Microwell holder.
- (7) 1 white dropper bottle of 6 ml Substrate/Chromagen.
- (8) 1 yellow or orange dropper bottle of Stop reagent.

b. Materials Required but not Provided:

- (1) Methanol - ACS grade or better.
- (2) Deionized or Distilled Water.

- (3) 250 ml graduated cylinder.
- (4) 125 ml container.
- (5) Filtering syringe (JM1000).
- (6) Sample collection tubes.
- (7) Waring high-speed blender with a one liter jar, or equivalent.
- (8) Sample grinder.
- (9) Balance.
- (10) Biotek EL 301 or an Awareness Technology Inc. Stat-Fax Model 303 Plus Microwell reader equipped with a 450-nm filter.
- (11) Eppendorf Repipettor, or equivalent, and 2.5 ml syringes.
- (12) 50 µl and 1000 µl Pipettor and pipette tips.
- (13) Paper towels, Kaydry paper or equivalent absorbent material.
- (14) Waste receptacle.
- (15) Timer: 3 channel minimum.
- (16) Waterproof marker, Sharpie or equivalent.
- (17) Wash bottle.
- (18) Deionized or distilled water.

6.10 STORAGE CONDITIONS

a. Storage Conditions.

- (1) The reagents supplied with the test kit can be used until the expiration date on the kit label when stored refrigerated at temperatures between 35° F and 46° F. **(DO NOT FREEZE)**
- (2) Return any unused microwells to their original foil bag and reseal them together with the desiccant provided.

- (3) The substrate/chromogen solution is light sensitive, therefore, avoid exposure to direct light.

b. Indication of Instability or Deterioration of Reagents.

- (1) Any bluish coloration of the red stained substrate/chromogen solution is indicative for deterioration and the reagent should be discarded.
- (2) A value of less than 0.6 absorbance units for the zero standard may indicate deterioration of reagents.

CHAPTER 12

RIDASCREEN® FAST AFLATOXIN SC TEST KIT

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12.1 GENERAL INFORMATION

The RIDASCREEN® FAST Aflatoxin SC test is a competitive enzyme immunoassay for the quantitative analysis of aflatoxin in select grains and commodities. **The test kit is limited to providing aflatoxin measurements between 5 – 100 ppb.**

12.2 PREPARATION OF SOLUTIONS

a. Extraction Solution.

The extraction solvent used in the RIDASCREEN® FAST Aflatoxin SC test is a methanol/water (distilled or deionized) mixture consisting of 70 percent methanol (ACS grade or better) and 30 percent water.

- (1) Using a graduated cylinder, measure 700 ml of methanol and place it into a clean carboy with spigot.
- (2) Add 300 ml deionized or distilled water to the methanol and shake vigorously until it is completely mixed.
- (3) Label the container stating the mixture (70 percent methanol and 30 percent water), date of preparation, and initials of technician who prepared the solution.
- (4) Store this solution at room temperature in a tightly closed container until needed.

NOTE: To prepare smaller or larger amounts of solution use the ratio of 7 parts methanol to 3 parts of deionized or distilled water.

b. Wash Solution.

- (1) Dissolve the contents of the packet containing the buffer salt in 1 liter of distilled water.
- (2) Swirl to mix.
- (3) Store this solution in a refrigerator until needed. The solution expires 4 weeks after preparation.

12.3 EXTRACTION PROCEDURES

- a. Transfer 50 grams of ground sample into an extraction mixing jar.
- b. Add 250 ml of the (70/30) methanol/water extraction solvent.
- c. Cover the extraction jar and blend on high speed for 2 minutes.
- d. Filter approximately 1.5 ml of the extract through a filtering syringe.
- e. Proceed to test procedures.

12.4 TEST PROCEDURES

- a. Sample Analysis.
 - (1) Allow reagents, microwells, and sample extracts to reach room temperature prior to running the test.
 - (2) Insert a sufficient number of wells into the microwell holder for all standards and samples to be tested. (For example: to test 7 samples use 8 wells - 1 for the standard and 7 for the test samples).

Test Strip

Well #	1	2	3	4	5	6	7	8
Sample	C 0	S1	S2	S3	S4	S5	S6	S7

Where C 0 is the zero control, S1 is sample 1, S2 is sample 2, S3 is sample 3, etc.

NOTE: Do not run more than 3 strips (23 samples) per set of control standards.

- (3) Using a new pipette tip for each standard and sample, pipet 50 µl of standard and prepared sample to separate wells.
- (4) Add 50 µl of enzyme conjugate (red capped bottle) into each well.
- (5) Add 50 µl of anti-aflatoxin antibody (black capped bottle) into each well.

- (6) Mix thoroughly by gently sliding the plate back and forth on a flat surface.
- (7) Incubate for 10 minutes (± 1.0 minutes) at room temperature (64 – 86° F).
- (8) Dump the contents of the wells. Turn the wells upside down and tap out on a paper towel until the remaining liquid has been removed.
- (9) Using a wash bottle, fill each well with distilled or deionized water or washing buffer solution. Empty the wells again and remove all remaining liquid. Repeat this step 2 times (total of 3 washes).
- (10) Add 100 μ l of substrate/chromagen (white dropper bottle) to each well.
- (11) Mix thoroughly by gently sliding the plate back and forth on a flat surface.
- (12) Incubate for 5 minutes (± 0.5 minutes) at room temperature (64 – 86° F). Cover the wells with a paper towel to protect them from light sources.
- (13) Add 100 μ l of stop solution (yellow or orange dropper bottle) to each well.
- (14) Mix thoroughly by gently sliding the plate back and forth on a flat surface.
- (15) Measure absorbance at 450 nm using the Biotek EL 301, or Awareness Technology Stat-Fax Model 303 PLUS microwell readers.

(Results must be read within 10 minutes)

b. Reading Results with the Microwell Reader.

- (1) Biotek EL 301 Microwell Reader.
 - (a) Make sure that the microwell reader is on and allowed to warm-up for a minimum of 15 minutes before using.
 - (b) Remove sample carriage and hit "Enter."
 - (c) Insert W2 filter and hit "Enter."
 - (d) Insert W1 filter (450 nm) and hit "Enter."

- (e) Hit "Clear" and then "Blank." This will cause the instrument to read air as the blank sample.
- (f) Load microwells into sample carriage so that the first control labeled 0 is in position A1.
- (g) Load the sample carriage into the strip reader so that position A1 is under the light beam of the reader.
- (h) Press "Read" and an absorbance value for A1 should appear in the display on the microwell reader. Record the value.
- (i) Slide the carriage to position A2 and press "Read." An absorbance value for A2 will appear. Record the value.
- (j) Repeat step (i) until absorbance values have been obtained for the control and all samples. Record the values.
- (k) Use the RIDA®SOFT Win Data software provided by r-Biopharm to convert the absorbance values into concentration values.

(2) Stat-Fax Model 303 PLUS Microwell Reader

- (a) To begin from the "Ready" prompt, press Menu, key in the test number, and then press Enter.
- (b) The screen will read, "Set carrier to A, press enter." Place the wells all the way to the right in the carrier. Push the carrier all the way to the left to line up the notch with the wells, then press enter. The carrier will advance into the reader, and it should start to print.
- (c) When the reader is finished reading the strip, the screen will read, "Plot Curve Y/N?"

Press "Yes" (1/A) to print the graph,

Press "No" (0) to skip this feature.
- (d) The screen will read, "Accept Curve Y/N ?"

Press "Yes" (1/A) to accept the curve and proceed to read another strip. When finished reading the second strip, press "Clear" twice and the results strip will print, "Test Ended."

Press "No" (0) to end the test.

12.5 REPORTING AND CERTIFYING TEST RESULTS

- a. Report all results on the pan ticket and the inspection log to the nearest whole ppb.
- b. Sample results over 100 ppb are reported as >100 ppb unless a supplemental analysis is performed.
- c. Refer to the Certification section of the handbook for more detailed certification procedures.

12.6 SUPPLEMENTAL ANALYSIS

- a. Diluting the Sample Extract.

If quantitative results are above the testing limits (i.e., 100 ppb) of the test kit, test results are reported as exceeding the limit. To determine and report an aflatoxin level higher than 100 ppb, the sample extract must be diluted so that a value between 5 and 100 ppb is obtained.

The final aflatoxin concentration is calculated by multiplying the results with the diluted extract by the dilution factor.

- b. Example.

If the original analysis reported the aflatoxin value at 130 ppb, the sample extract would be diluted using the following procedures in order to obtain a true value.

- (1) Dilute 5 ml of the original extract (obtained from step c., section 6.3) with 5 ml of the extraction solvent mixture. The total volume is 10 ml. This is a 1 to 2 dilution (compares volume in the beginning with the total volume in the end).
- (2) Multiply the analytical results obtained by 2 to obtain the actual aflatoxin concentration. For example, if 74 ppb was the original value obtained with the diluted extract, the actual concentration in the original sample was 148 ppb.

$$\text{True Aflatoxin Value} = \frac{\text{Total Volume}}{\text{Initial Extract Volume}} \times \text{Aflatoxin Result}$$

$$\begin{aligned}\text{True Aflatoxin Value} &= (10 \div 5) \times 74 \text{ ppb} \\ &= 2 \times 74 \text{ ppb} = 148 \text{ ppb}\end{aligned}$$

12.7 CLEANING LABWARE

a. Negative Tests (≤ 20 ppb).

(1) Labware.

Prepare a solution consisting of dishwashing liquid and water. Completely submerge the used glassware, beakers, etc., wash thoroughly, then rinse with clean water before reusing.

(2) Disposable Materials.

Place materials in a garbage bag for routine trash disposal.

b. Positive Tests (> 20 ppb).

(1) Labware.

Prepare a bleach solution consisting of 1 part bleach to 10 parts water (e.g., 100 ml bleach to 1,000 ml water). Completely submerge the used glassware, beakers, etc., and soak for at least 5 minutes. Remove items from the bleach/water solution, submerge in a dishwashing liquid/water solution, wash thoroughly, then rinse with clean water before reusing.

(2) Disposable Materials.

Prepare a bleach solution consisting of 1 part bleach to 10 parts water in a plastic pail labeled "bleach solution". Soak disposable materials, such as used columns, cuvettes, vials, test kit components, etc., for at least 5 minutes. Pour the liquid down the drain and place the materials in a garbage bag and discard.

12.8 WASTE DISPOSAL

a. Negative Results (≤ 20 ppb).

If the test result is negative (equal to or less than 20 ppb), discard the syringe into a plastic garbage bag for disposal.

b. Positive Results (> 20 ppb).

If the result is positive (more than 20 ppb), the remaining ground portion must be decontaminated, using bleach, prior to disposal. Discard the filter syringe and remaining ground portion into a plastic garbage bag for disposal. The bleach rinse filtrate collected may be treated as a non-hazardous solution and disposed of by pouring down the drain.

12.9 EQUIPMENT AND SUPPLIES

a. Materials Supplied in Test Kits

- (1) 1 microtiter plate.
- (2) 48 antibody coated microwells.
- (3) 1 aflatoxin standard solution of 1.3 ml of 0 ppb aflatoxins.
- (4) 1 red-capped bottle of 3 ml peroxidase conjugated aflatoxin solution.
- (5) 1 black-capped bottle of 3 ml anti-aflatoxin antibody.
- (6) 1 white dropper bottle of 6 ml Substrate/Chromagen.
- (7) 1 yellow or orange dropper bottle of Stop reagent.
- (8) 1 washing buffer.

b. Materials Required but not Provided:

- (1) Methanol - ACS grade or better.
- (2) Deionized or Distilled Water.

- (3) 250 ml graduated cylinder.
- (4) 125 ml container.
- (5) Filtering syringe (JM1000), Whatman No. 1 filter paper, or equivalent.
- (6) Sample collection tubes.
- (7) Waring high-speed blender with a one liter jar, or equivalent.
- (8) Sample grinder.
- (9) Balance.
- (10) Biotek EL 301 or an Awareness Technology Inc. Stat-Fax Model 303 Plus Microwell reader equipped with a 450-nm filter.
- (11) Eppendorf Repipettor, or equivalent, and 2.5 ml syringes.
- (12) 50 µl, 100 µl, and 1000 µl pipettor and pipette tips.
- (13) Paper towels, Kaydry paper or equivalent absorbent material.
- (14) Waste receptacle.
- (15) Timer: 3 channel minimum.
- (16) Waterproof marker, Sharpie or equivalent.
- (17) Wash bottle.

12.10 STORAGE CONDITIONS

a. Storage Conditions.

- (1) The reagents supplied with the test kit can be used until the expiration date on the kit label when stored refrigerated at temperatures between 35° F and 46° F. **(DO NOT FREEZE)**
- (2) Return any unused microwells to their original foil bag and reseal them together with the desiccant provided.

- (3) The substrate/chromogen solution is light sensitive, therefore, avoid exposure to direct light.

b. Indication of Instability or Deterioration of Reagents.

- (1) Any bluish coloration of the red stained substrate/chromogen solution is indicative for deterioration and the reagent should be discarded.
- (2) A value of less than 0.6 absorbance units for the zero standard may indicate deterioration of reagents.

CHAPTER 13

QUICKTOX TEST KIT

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13.1 GENERAL INFORMATION

The QuickTox test kit uses lateral flow test strip technology that provides qualitative (equal to or less than a specified threshold) results.

13.2 PREPARATION OF EXTRACTION SOLUTION

The extraction solvent used in the QuickTox test method is a methanol/water (distilled or deionized) mixture consisting of 70 percent methanol (Reagent grade or better) and 30 percent water.

- a. Using a graduated cylinder, measure 700 ml of methanol and place it into a clean carboy with spigot.
- b. Add 300 ml deionized or distilled water to the methanol and shake vigorously until it is completely mixed.
- c. Label the container stating the mixture (70 percent methanol and 30 percent water), date of preparation, and initials of technician who prepared the solution.
- d. Store this solution at room temperature in a tightly closed container until needed.

NOTE: To prepare smaller or larger amounts of solution use the ratio of 7 parts methanol to 3 parts of deionized or distilled water.

13.3 EXTRACTION PROCEDURES

- a. Transfer 50 grams of ground sample into an extraction mixing jar.
- b. Add 100 ml of the (70/30) methanol/water extraction solvent.
- c. Cover the extraction jar and shake by hand for 2 minutes. If a mechanical shaker is used shaking time may be reduced to 1 minute.
- d. After shaking, the sample will immediately begin to separate into 2 layers. The top (yellowish) layer containing the aflatoxin residues will be used for testing.

13.4 TEST PROCEDURES

a. Reaction Vial.

- (1) Using the fixed volume pipette included in the test kit, place 150 microliters (150 μ l) **tap** water into a reaction vial.
- (2) Using the same fixed volume pipette, remove 150 μ l from the top (yellowish) layer of the extract. Add the extraction solution to the reaction vial containing water.
- (3) Mix water and sample extraction solution by stirring with the tip of the fixed volume pipette.

NOTE: To ensure correct volumes are used to prepare the test sample, a fixed volume pipette is included with the kit. When a liquid drawn to the top of the straw end of the pipette is dispensed, 150 μ l will be expelled into the reaction vial. Any overfill is retained in the pipette. After diluting the sample the final volume in the reaction vial should be 300 μ l. Do not reuse diluted samples. Use a new fixed volume pipette and reaction vial for each sample.

b. Test Strips.

- (1) Allow refrigerated canisters to come to room temperature before opening. Remove the QuickTox strips to be used then immediately reseal the canister. Avoid bending the strips.
- (2) Place the strip into the reaction vial containing the diluted sample extract. The arrow tape on the end of the strip should point into the reaction vial.
- (3) The sample extract will travel up the strip. Reaction vials will stand on their own or may be inserted into the cardboard racks provided.
- (4) Allow the strip to develop for 5 minutes before making final assay interpretations. Negative sample results may become obvious more quickly (2 – 3 minutes).

c. Interpreting the Lateral Flow Test Strip.

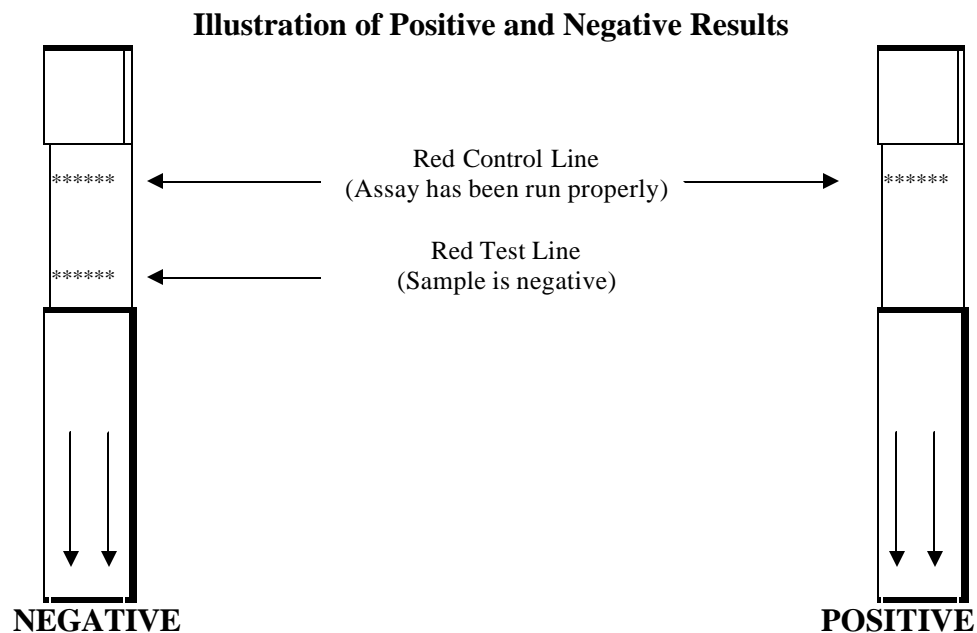
Development of a Control Line within 5 minutes indicates that the strip has functioned properly. Any strip that does not develop a Control Line should be discarded. A second preparation of the extract (using a fresh 1:2 dilution) should be made and tested using another strip.

(1) Negative Result.

A sample containing aflatoxin residues less than or equal to 20 ppb will develop 2 distinct lines, the Control Line and the Test Line, in the test area.

(2) Positive Result.

A sample containing aflatoxin residues in excess of 20 ppb will develop 1 distinct line, the Control Line.



13.5 REPORTING AND CERTIFYING TEST RESULTS

- a. Report results on the pan ticket and inspection log as being equal to or less than 20 ppb (≤ 20 ppb), or as exceeding 20 ppb (> 20 ppb), as applicable.
- b. Certify results as being equal to or less than 20 ppb or exceeding 20 ppb, as applicable.
- c. Refer to the Certification section of the handbook for more detailed certification procedures.

13.6 CLEANING LABWARE

a. Negative Tests (≤ 20 ppb).

(1) Labware.

Prepare a solution consisting of dishwashing liquid and water. Completely submerge the used extraction mixing jars, wash thoroughly, then rinse with clean water before reusing.

(2) Disposable Materials.

Place materials in a garbage bag for routine trash disposal.

b. Positive Tests (> 20 ppb).

(1) Labware.

Prepare a bleach solution consisting of 1 part bleach to 10 parts water (e.g., 100 ml bleach to 1,000 ml water). Completely submerge the used extraction mixing jars and soak for at least 5 minutes. Remove items from the bleach/water solution, submerge in a dishwashing liquid/water solution, wash thoroughly, then rinse with clean water before reusing.

(2) Disposable Materials.

Prepare a bleach solution consisting of 1 part bleach to 10 parts water in a plastic pail labeled "bleach solution". Soak disposable materials, such as used test strips and pipettes, for at least 5 minutes.

Pour off the liquid down the drain and place the materials in a garbage bag and discard.

13.7 WASTE DISPOSAL

a. Negative Results (≤ 20 ppb).

If the test result is negative (equal to or less than 20 ppb), dispose of any remaining liquid filtrate in the chemical waste container. Discard the sample slurry (ground material) into a plastic garbage bag for disposal.

b. Positive Results (> 20 ppb).

If the result is positive (more than 20 ppb), the slurry (ground portion) remaining in the sample extraction jar must be decontaminated prior to disposal. After disposing of the remaining filtered extract in the chemical waste container, pour approximately 50 ml of bleach solution into the sample extraction jar and shake to mix with the sample slurry. After the slurry and bleach solution separate, handle the bleach rinse filtrate as a non-hazardous solution and dispose of by pouring the liquid down the drain. Discard the sample slurry (ground portion) paper into a plastic garbage bag for disposal.

13.8 EQUIPMENT AND SUPPLIES

a. Materials Supplied in Test Kits

- (1) 50 QuickTox strips packed in a moisture-resistant container.
- (2) 50 fixed volume transfer pipettes.
- (3) 50 reaction vials.

b. Materials Required but not Provided:

- (1) Timer (5 minute capacity).
- (2) Felt tipped pens.
- (3) Balance.

- (4) Sample Grinder.
- (5) Methanol - Reagent grade or better.
- (6) Deionized or Distilled water.
- (7) Sample extraction jars.
- (8) Orbital/rotary shaker.
- (9) Tap water.

13.9 STORAGE CONDITIONS

a. Storage Conditions.

Test kits should be refrigerated between 36° - 48°F.

b. Precautions.

- (1) Do not use the test kits beyond the noted expiration date.
- (2) Prolonged exposure to high temperatures may adversely affect the test results.
- (3) Do not open the desiccated canister until ready to use the strips.